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## Stereoselective synthesis of the $C_{14}$ - $C_{26}$ fragment of the cytotoxic macrolide FD-891

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**Abstract**—A stereoselective synthesis of the  $C_{14}$ — $C_{26}$  fragment of the naturally occurring, cytotoxic macrolide FD-891, is described. Asymmetric Evans aldol reactions and aldehyde Brown allylations are key steps of the synthesis. © 2004 Elsevier Ltd. All rights reserved.

The cytotoxic metabolite FD-891 was isolated from the fermentation broth of *Streptomyces graminofaciens* A-8890 and was found active against several tumor cell lines. In addition, it was found to potently prevent both perforin- and FasL-dependent CTL-mediated killing pathways. In contrast to the structurally related concanamycin A, however, it was unable to inhibit vacuolar acidification.<sup>1</sup> We have now faced the problem of synthesizing this bioactive metabolite and have chosen the convergent retrosynthesis shown in Scheme 1. According to it, the molecule of FD-891 is disconnected to frag-



Scheme 1. Structure of FD-891 and main retrosynthetic disconnection.

*Keywords*: Macrolides; Cytotoxicity; FD-891; Asymmetric allylboration; Boron aldol reactions.

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ments A (C<sub>1</sub>–C<sub>13</sub>) and B (C<sub>14</sub>–C<sub>26</sub>) via an esterification and a Heck coupling. Which one of these two reactions will be the final macrocyclization process still remains an open issue to be decided at a later stage. Indeed, both macrolactonizations<sup>2</sup> and intramolecular Heck reactions<sup>3</sup> are amply represented in the literature.

In the present communication, we will describe the synthetic work performed to achieve the preparation of fragment **B**, which carries the protecting groups R = TBS (*t*-butyldimethylsilyl) and MOM (methoxymethyl).<sup>4</sup> This fragment, which contains 7 of the 12 stereocenters of the molecule, was further retrosynthetically disconnected as shown in Scheme 2. One key structural transformation in this retrosynthesis ( $B \rightarrow I$ ) is the



Scheme 2. Retrosynthetic disconnection of fragment B.

stereoselective allylation of a chiral  $\alpha$ -methyl aldehyde and was planned to be performed in an asymmetric way using one of Brown's chiral allylboration reagents.<sup>5</sup> The protecting group TPS (*t*-butyldiphenylsilyl) was selected with the idea in mind of its later selective cleavage in the presence of two TBS groups. The two other key retrotransformations,  $\mathbf{II} \rightarrow \mathbf{III}$  and  $\mathbf{III} \rightarrow \mathbf{IV}$ , are aldol reactions conceived to create the C<sub>22</sub>-C<sub>25</sub> dipropionate



Scheme 3. Stereoselective synthesis of compound B. Reagents and conditions: (a) (COCl)<sub>2</sub>, DMSO, CH<sub>2</sub>Cl<sub>2</sub>, Et<sub>3</sub>N, 1h, 0°C; (b) 16, Bu<sub>2</sub>BOTf, Et<sub>3</sub>N, 0°C, then 3, 2.5h, 0°C, 89% overall from 2; (c) MeNHOMe, AlMe3, THF, 3h, rt, 79%; (d) TBSOTf, 2,6-lutidine, CH<sub>2</sub>Cl<sub>2</sub>, rt, 1h, 92%; (e) DIBAL, THF, -78°C, 30min; (f) 17, Bu<sub>2</sub>BOTf, Et<sub>3</sub>N, 0°C, then 6, 3h, 0°C, 75% overall from 5; (g) TBSOTf, 2,6-lutidine, CH<sub>2</sub>Cl<sub>2</sub>, rt, 1 h, 90%; (h) H<sub>2</sub>O<sub>2</sub>, aq LiOH, THF,  $0^{\circ}C \rightarrow rt$ , overnight; (i) MeNHOMe, 1,1'-carbonyldiimidazole, CH<sub>2</sub>Cl<sub>2</sub>, rt, 12h, 80% overall from 8; (j) MeMgBr, THF, 0°C, 1h, 70%; (k) Me<sub>2</sub>AlCl (2.5 equiv), Bu<sub>3</sub>SnH, CH<sub>2</sub>Cl<sub>2</sub>, -90 °C, 1 h, 91% (92:8 diastereoisomeric mixture); (1) MeOTf, 2,6-di-t-butylpyridine, CHCl<sub>3</sub>, Δ, 4h, 84%; (m) 10% NaOH, MeOH, Δ, 30h, 84%; (n) (COCl)<sub>2</sub>, DMSO, CH<sub>2</sub>Cl<sub>2</sub>, Et<sub>3</sub>N, 20min, 0°C; (o) allylBIpc<sub>2</sub> [from (-)-DIP-Cl and allylmagnesium bromide], Et<sub>2</sub>O, 1h, -90°C, 55% overall from 13 as a single stereoisomer; (p) MOMCl, EtNi-Pr2, CH2Cl2, rt, overnight, 79%.

segment. In the actual synthesis, both aldol steps were executed with the aid of the chiral oxazolidinones developed by Evans and his group.<sup>6</sup> The ultimate chirality source was the commercially available ester **1**.

Scheme 3 depicts the actual synthetic sequence, which led to compound **B**. The chiral, commercially available ester 1 was converted into the known primary alcohol  $2^7$  via a literature procedure. Swern oxidation of the latter to aldehyde 3 was followed by Evans asymmetric aldolization using the Z boron enolate of the chiral oxazolidinone 16.6a This provided aldol adduct 4 as an essentially single stereoisomer. Conversion of 4 into the Weinreb amide<sup>8</sup> and silvlation afforded 5, which was then reduced (DIBAL) to aldehyde 6. The latter compound was submitted to a second aldolization with Evans oxazolidinone 17,<sup>6a</sup> followed by silulation and amidation. This yielded Weinreb amide  $9,^9$  which was converted into methyl ketone 10 by treatment with methylmagnesium bromide. Stereoselective reduction of the carbonyl group of **10** under chelation control<sup>10-13</sup> to alcohol 11 and subsequent O-methylation with methyl triflate/2,6-di-tert-butylpyridine<sup>14</sup> afforded compound 12. Selective cleavage of the OTPS group in 12 under alkaline conditions<sup>15</sup> provided the primary alcohol 13, which was then oxidized to aldehyde 14. Asymmetric allylation of the latter<sup>5</sup> afforded secondary alcohol 15, which, through protection of the secondary alcohol group as its MOM derivative,16 afforded the desired fragment **B**.<sup>17</sup>

In summary, a stereoselective synthesis of the  $C_{14}$ – $C_{26}$  fragment of the cytotoxic macrolide FD-891 has been achieved. Studies toward the total synthesis of the natural product are underway and will be published in due course.

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19 ppm. This indicates that it is the acetonide of a *syn*-1, 3-diol<sup>13</sup> and can only be that indicated below, as the internal acetonide would necessarily be *anti*.



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- 17. Oil;  $[\alpha]_D = -3.1$  (*c* 0.86, CHCl<sub>3</sub>), <sup>1</sup>H NMR (500 MHz)  $\delta$  5.82 (1H, m, H-2), 5.10 (1H, dd, J = 17, 1.5 Hz, H-1t), 5.04 (1H, H-1t), 5.04 (1H, H-1t), 5.04 (1H, H-1t))dd, J = 10, 1 Hz, H-1c), 4.65 (2H, AB system, J = 7 Hz, MOM), 3.81 (1H, dd, J = 5.8, 1.6 Hz, H-10), 3.70 (1H, m, H-8), 3.47 (1H, m, H-4), 3.37 (3H, s, OMe), 3.30 (3H, s, OMe), 3.14 (1H, quint, J = 6.5 Hz, H-12), 2.30 (2H, m, H-3), 1.75-1.50 (5H, br m, H-5/H-6/H-9), 1.45-1.40 (2H, m, H-7), 1.12 (3H, d, *J* = 6.5 Hz, H-13), 0.94 (3H, d, *J* = 7 Hz, Me-C), 0.91 (3H, d, J = 7 Hz, Me-C), 0.90 (9H, s, t-Bu), 0.89 (9H, s, t-Bu), 0.86 (3H, d, J = 7 Hz, Me-C), 0.09 (3H, s, Me-Si), 0.07 (3H, s, Me-Si), 0.06 (3H, s, Me-Si), 0.05 (3H, s, Me-Si).  $^{13}$ C NMR (125 MHz)  $\delta$  18.5, 18.3 (C), 135.6, 81.4, 79.9, 73.4, 72.6, 43.4, 41.2, 36.6 (CH), 116.7, 96.1, 36.0, 33.4, 27.2 (CH<sub>2</sub>), 56.4, 55.6, 26.1 (×3), 26.0 (×3), 16.5, 14.6, 11.1, 10.6, -3.3, -3.5, -4.0, -4.1 (CH<sub>3</sub>). HR EIMS m/z (% rel. int.) 517.3756 (M<sup>+</sup> – t-Bu, 3), 283 (20), 231 (55), 139 (34), 59 (100). Calcd for C<sub>31</sub>H<sub>66</sub>O<sub>5</sub>Si<sub>2</sub>-t-Bu, 517.3744.